

THE BASIC GLUCOSIDES RELATED TO THE
BIOSYNTHESIS OF INDOLE AND IPECAC ALKALOIDS

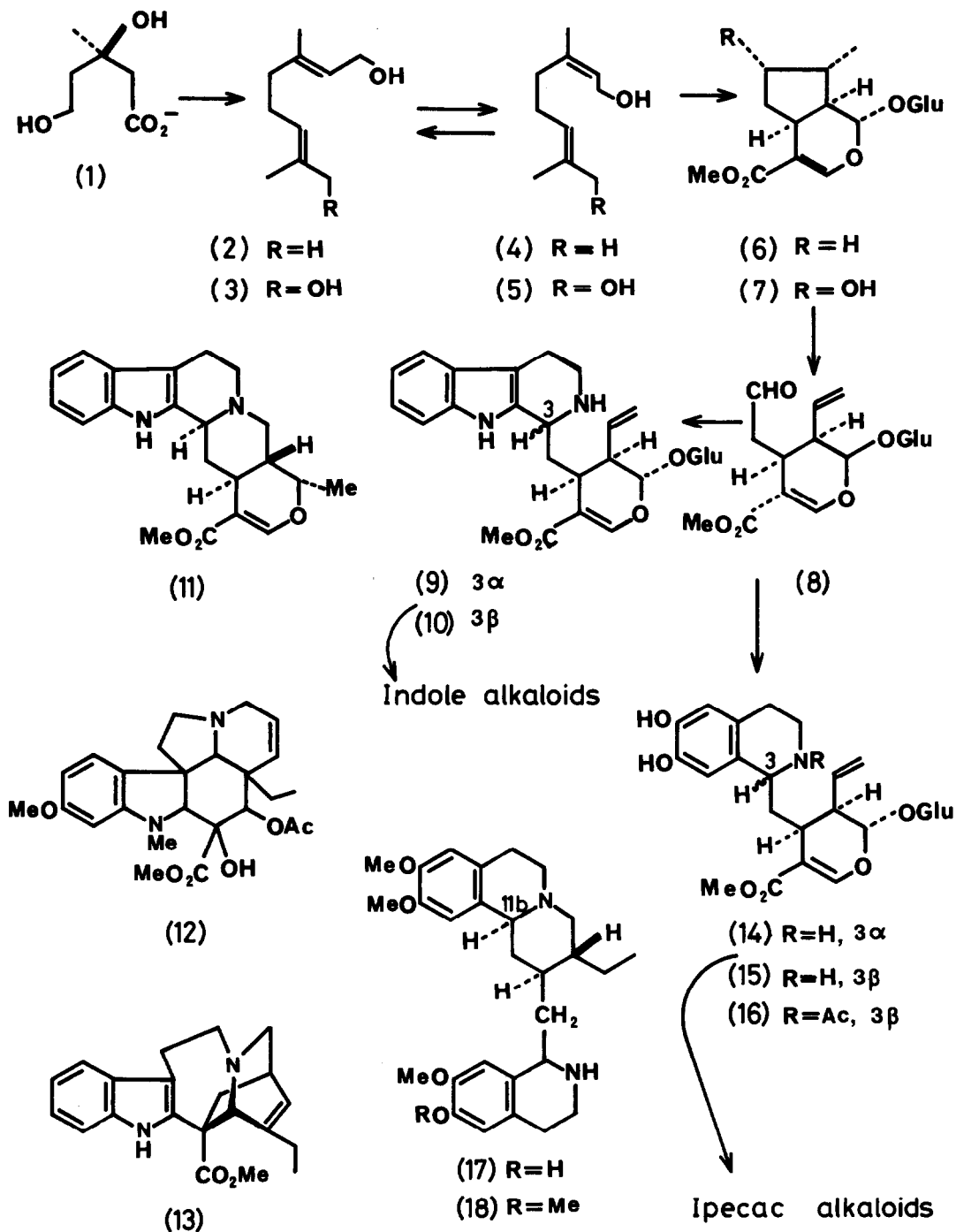
Alan R. Battersby*, Norman G. Lewis
and James M. Tippet

University Chemical Laboratory, Lensfield Road,
Cambridge, CB2 1EW.

During the nineteen sixties, the biosynthetic pathways to the indole and Ipecac alkaloids were largely elucidated¹⁻³ by the efforts of several research groups. The non-tryptamine portion of the indole alkaloids was found to be derived from mevalonate (1) via geraniol (2) and nerol (4) which after hydroxylation to (3) and (5) yielded deoxyloganin (6) and subsequently loganin (7). A fascinating cleavage step then generated secologanin (8) which with tryptamine gave rise to strictosidine⁴ (9) [\equiv isovincoside⁵] together with vincoside⁵ (10). The experimental approach for these studies involved tracer experiments in vivo combined with isolation work and structure determinations. As a result, many new substances were discovered among them being the biosynthetic intermediates deoxyloganin⁶ (6), secologanin⁷ (8) and the basic glucosides (9) and (10), these four having been predicted¹ to be involved with biosynthesis of indole alkaloids.

Our early tracer studies⁵ with a mixture of the basic glucosides (9) and (10) established that one or both could be specifically incorporated by Catharanthus roseus (\equiv vinca rosea) plants into representatives of the three major indole alkaloid families, viz. ajmalicine (11), vindoline (12) and catharanthine (13). The glucosides were doubly-labelled at well separated sites and the isolated alkaloids were suitably degraded to establish incorporation without randomisation. In addition, (9) & (10) were both isolated from C. roseus plants.⁵ Feedings of the separated [O-methyl-³H] bases (9) and (10) to the plants⁵ led to the conclusion that 3 β -isomer (10) was on the biosynthetic pathway to the alkaloids rather than the 3 α -isomer (9) and other workers⁸ also found no appreciable incorporation of labelled (9) into indole alkaloids when it was administered to C. roseus plants. However, the above double-labelling experiments established the main relationship of the indolic glucoside system with the alkaloids and the lack of match between the configuration at C-3 of (10) and the corresponding site of ajmalicine (11) remained as an incongruous spot on the large biosynthetic canvas.

Recent experiments⁹ confirmed the foregoing double-labelling results and, satisfyingly, the 3 α -isomer (9) was found to be incorporated into the alkaloids. However, (9) was again not incorporated under certain feeding conditions due to



Scheme. Biosynthesis of Indole and Ipecac Alkaloids

degradation of the precursor by non-specific glucosides and this is almost certainly one of the factors in the variable incorporation results observed in the basic glucoside area. We have therefore re-examined this stereochemical point using samples of (9) & (10) synthesised⁷ from [aryl-³H]tryptamine and secologanin (8), taking advantage of the advances in fractionation methods over the past 10 years. The results (Table) confirm that the 3 α -isomer[†] (9) is incorporated, rather than the 3 β (10) and additional confirmation¹⁰ has been adduced.

In the Ipecac series (see Scheme), the earlier work had largely defined the pathway³ and it followed the same track as for the indole alkaloids, outlined above, up to secologanin (8). This then condensed with dopamine, rather than with tryptamine, to form desacetylisoipecoside (14) and desacetylipecoside (15). Fresh syntheses of these two glucosides have been carried out using [1-¹⁴C]dopamine and secologanin (8) and the separated products were fed

TABLE Tracer experiments on Vinca rosea and Cephaelis ipecacuanha

Plant and isolated alkaloids	Precursor and Incorporations (%)			
	3 α -Isomer (9)	3 β -Isomer (10)	3 α -Isomer (14)	3 β -Isomer (15)
<u>V. rosea</u>				
Ajmalicine (11)	1.1	<0.01	-	-
Vindoline (12)	0.96	<0.01	-	-
Catharanthine (13)	1.5	<0.01	-	-
<u>C. ipecacuanha</u>				
Cephaeline (17)	-	-	1.6; 1.1	<0.003; <0.01
Emetine (18)	-	-	0.32; 0.28	<0.001; <0.001
Ipecoside ¹¹ (16)	-	-	<0.025; <0.011	0.32; 0.90

individually to Cephaelis ipecacuanha plants. The results in the Table clearly show that the 3 α -isomer, desacetylisoipecoside (14) is the precursor of cephaeline (17) and emetine (18), not³ the 3 β -isomer. The 3 α configuration of desacetylisoipecoside (14) matches¹¹ that of the corresponding centre (C-11b) of cephaeline (17) and emetine (18) and so here too the biosynthetic relationship is now stereochemically unexceptional.

[†] To simplify the literature, it is best that (9) should be known as strictosidine⁴ and (10) as vincoside.

Acknowledgements

We thank Dr. I. Kompis (F. Hoffmann-La Roche, Basle) for a gift of secologanin, the N.R.C. (Canada) for a postdoctoral award (to N.G.L.) and the Nuffield Foundation and S.R.C. for financial support.

REFERENCES

1. A.R. Battersby, Pure and Appl. Chem., 1967, 14, 117.
2. Reviewed by (a) R.J. Parry, in "The Catharanthus Alkaloids", eds. W.I. Taylor & N.R. Farnsworth, Dekker, New York 1975, p. 141 (b) G.A. Cordell, Lloydia, 1974, 37, 219 (c) A.R. Battersby, Spec. Period. Rep. Chem. Soc., The Alkaloids, 1971, 1, 31.
3. A.R. Battersby & R.J. Parry, Chem. Comm., 1971, 901 and refs. therein.
4. G.N. Smith, Chem. Comm., 1968, 912; K.T.D. De Silva, G.N. Smith & K.E.H. Warren, Chem. Comm., 1971, 906.
5. A.R. Battersby, A.R. Burnett, & P.G. Parsons, J. Chem. Soc. (C), 1969, 1193.
6. H. Inouye, S. Ueda, Y. Aoki, & Y. Takeda, Tetrahedron Letters, 1969, 2351; A.R. Battersby, A.R. Burnett, & P.G. Parsons, Chem. Comm., 1970, 826.
7. A.R. Battersby, A.R. Burnett, & P.G. Parsons, J. Chem. Soc. (C), 1969, 1187.
8. R.T. Brown, G.N. Smith, & K.S. Stapleford, reported in ref. 2(b), p. 236.
9. J. Stöckigt and M.H. Zenk, J.C.S. Chem. Comm., 1977, 646.
10. A.I. Scott, S.L. Lee, P. de Capite, M.G. Culver, and C.R. Hutchinson, Heterocycles, 1977, 7 (2), 979.
11. O. Kennard, P.J. Roberts, N.W. Isaacs, F.H. Allen, W.D.S. Motherwell, K.H. Gibson and A.R. Battersby, Chem. Comm., 1971, 899 and refs. therein.

(Received in UK 2 October 1978)